

Carbon fixation and partitioning in coffee seedlings infested with *Pratylenchus coffeae*

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Abstract

The objective was to study CO₂ fixation and photoassimilate partition in coffee (*Coffea arabica*) seedlings infested with the lesion nematode *Pratylenchus coffeae*. Seedlings infested with 0, 1000 and 8000 *Pratylenchus coffeae* nematodes were exposed to ¹⁴CO₂ and the incorporation and distribution of radioactivity were followed in the roots, stems and leaves. Fresh mass, pigments, soluble sugars, sucrose and specific radioactivity of sucrose in the plant parts were determined. At the highest level of infestation almost all the parameters were significantly changed showing the carbon fixation in the leaves and partitioning to the roots were decreased. Since lesion nematodes are not sedentary and do not form feeding sites that could be characterised as metabolic sinks, it is suggested that their damage is more readily expressed by the leaves, through a reduction in photosynthesis and phloem transport.

The root lesion nematode, *Pratylenchus coffeae*, (Zimmermann, 1898) Filipjev and Schuurmans Stekhoven, 1941 was first reported to infect coffee (*Coffea arabica*) by Monteiro and Lordello (1974). This nematode is a major pest in several countries where this crop is grown (Campos et al., 1990; Inomoto et al., 1998). Poor root growth, leaf shedding and nutritional deficiency are typical symptoms of infested coffee trees (Monteiro and Lordello, 1974). Reduction in photosynthesis also occurs (Kubo et al., 2003).

In view of some variation in terms of host-range variability (Edwards and Wehunt, 1973) and recent morphological and molecular studies showing differences among isolates (Duncan et al., 1999), the taxonomy of *Pratylenchus coffeae* has been re-examined. An isolate from *Citrus aurantium* was recently considered as a new species (Inserra et al., 2001). Two other Brazilian isolates, K₅ from *C. arabica* and M₂ from *Aglaonema* spp. are differ-

entiated by their host-range (Silva and Inomoto, 2002). The isolate K₅ is more pathogenic than M₂, causing almost complete destruction of the roots, and inducing chlorosis and leaf fall (Silva and Inomoto, 2002; Kubo et al., 2003).

Compared to *Meloidogyne* spp., very little is known about the damage caused by *Pratylenchus* in terms of growth and development of coffee (Kubo et al., 2003). Therefore, the aim of this work was to study the effect of *P. coffeae* on ¹⁴CO₂ assimilation and radioactivity partitioning in this crop, using isolate K₅ (Silva and Inomoto, 2002). Labelled CO₂ has long been used as a useful technique to study metabolic alterations in plants infested by nematodes (Bird and Loveys, 1975).

Nematodes maintained on alfalfa callus (Riedel et al., 1973) were extracted (juveniles and adults) from 45 to 90 day old cultures after the last transfer to new calli using the Baermann method, with modifications for flat recipient (Southey, 1986).

Five-month-old coffee (*C. arabica* cv. Catuaí Vermelho) seedlings, growing in plastic pots containing 450 ml of soil previously sterilised with methyl bromide (150 ml of CH₃Br/1000 l of soil), were inoculated with nematode suspension adjusted to obtain 1000 and 8000 nematodes/seedling. Control plants were not inoculated. The plants were kept under greenhouse conditions being watered every second day. After 45 days of inoculation, the plants were exposed to ¹⁴CO₂. Each pot containing one seedling was enclosed in a plastic bag together with two Eppendorf tubes inside, one with 0.5 ml 3 M HCl and other with 50 µCi NaH₃¹⁴CO₂ (58 mCi mmol⁻¹, Amersham-Pharmacia). The contents of the Eppendorfs were mixed by external handling and the seedlings were maintained in a glasshouse under a net cutting 25% of the natural light, from 8:00 a.m. to 13:00 p.m., when the bags were opened. The next morning (8:00 a.m.), leaves, stems and roots were collected and weighed. Roots were washed under tap water flow and blotted dry before weighing. The material was frozen at -60 °C before being extracted twice with methanol (2 × 10 ml/5 g) in a Polytron homogeniser. The debris was eliminated by filtration followed by centrifugation (10,000 rpm for 15 min) and 1 ml aliquots were used for the determination of radioactivity after addition of scintillation fluid. The same extracts were used for determination of pigments (Lichtenthaler and Wellburn, 1983), total soluble sugars (Dubois et al., 1956) and sucrose (Van Handel, 1968). Fractions were vacuum concentrated (SpeedVac, Savant) and applied on 3MM Whatmann paper for chromatographic separation of the soluble sugars (Chaplin, 1986). Chromatography was developed for 72 h and the sugars visualised by spraying with 5% H₂SO₄ and heating at 100 °C until colour development. Sucrose was identified by comparison with pure sucrose applied laterally on the paper. The identified spots were cut out from the chromatography paper and homogenised in the Polytron with 10 ml ethanol. After centrifugation, aliquots were taken for determination of the radioactivity. It was thereby possible to calculate the specific radioactivity for sucrose. The experiment was repeated twice, separated by a 1-week interval. The first assay was carried out with five replicates and the second with eight. In the first experiment the stems were not collected. Carbohydrate determinations were made only in the second assay.

Both assays showed that infection by the isolate K₅ of *P. coffeae* caused growth reduction in coffee seedlings (Figure 1A and D). Despite the simultaneous inoculation of the seedlings of both assays and a similar fresh mass accumulation (see Figure 1A and D), less radioactivity was detected in the second assay (Figure 1B and E). This probably occurred because in this assay the incubation with ¹⁴CO₂ was carried out on a cloudy day. However, data from both experiments showed a similar trend, that is, seedlings infected with 8000 nematodes assimilate less radioactivity indicating lower photosynthesis. These data are consistent with the radioactivity per mass data (Figure 1C and F) since there was a decrease as the inoculation nematode level was increased. In addition, the low radioactivity per root mass indicated that less sucrose was transported from the shoot to the roots in infested seedlings.

Stem radioactivity in the second assay was similar to the leaves (Figure 1F). A possible explanation is that the stem in young coffee plants has chlorophyll and, therefore, may be active in photosynthesis. A second possibility is the contribution of the radioactivity of the transported compounds in the xylem.

The data shown in Figure 1 demonstrate the contrast in terms of carbon partitioning between plants infected by lesion nematodes and sedentary nematodes. Carneiro et al. (2000) observed that soybean plants infected with *M. incognita* and *M. javanica* had higher total radioactivity and specific mass radioactivity in the roots, as the inoculum level increased. This was probably related with the formation of the feeding sites in the roots, acting as strong metabolic sinks. Since *Pratylenchus* is not sedentary and does not form feeding sites, the destruction of the root system by this nematode seems to be more readily felt by the leaves, leading to a faster decrease in carbon assimilation.

In addition to the decrease of mass accumulation and carbon fixation (Figure 1), chlorophyll in the leaves also decreased with infestation by *P. coffeae* (Figure 2). The inverse relationship between sucrose and total soluble sugars is of special interest (Figure 2B and C). The decrease of sucrose in the leaves with nematode infestation follows the reduction in carbon fixation, seen in Figure 1. On the other hand, the increase in soluble sugars might be well explained by starch hydrolysis due to a higher respiration rate since the

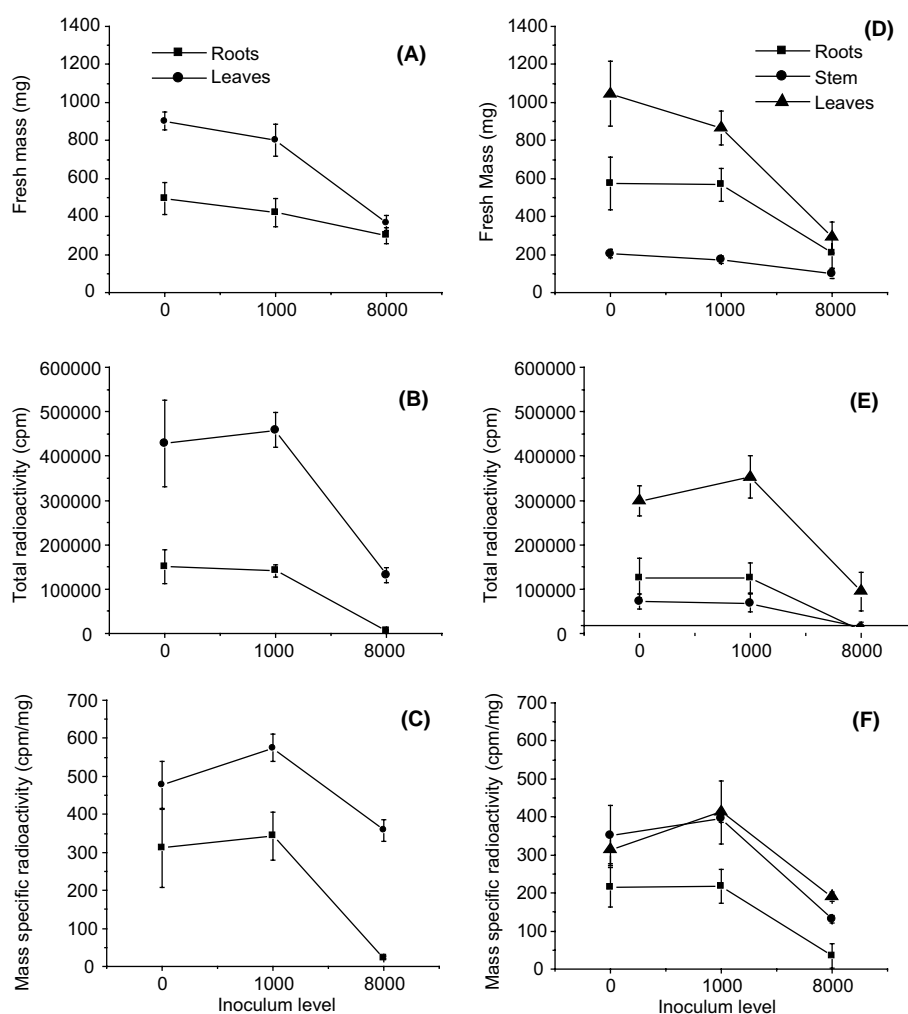


Figure 1. The effect of *P. coffeae* on fresh mass (A, D), total radioactivity distribution (B, E) specific mass radioactivity (C, F) in coffee seedlings of the assays 1 (A–C) and 2 (D–F). Bars indicated standard deviation. Means of five (assay 1) and eight (assay 2) replicates.

specific radioactivity of sucrose in the roots decreased (Figure 2D), clearly indicating reduced transport of sucrose from the leaves. In addition, the discrete reduction of endogenous sucrose in the roots (Figure 2C) might also be due to an increase of invertase activity as observed for diseased plants (Sturm, 1999) channelling the resulting reducing sugars for respiration. An increase in respiration by nematode infestation has been either suggested or reported (Hussey, 1985; Poskuta et al., 1986; Carneiro et al., 2000), although all results were obtained with root knot nematodes. The way lesion nematodes attack plants and the stress they

provoke might be comparable to mechanical injuries, which cause a significant rise in the respiratory metabolism (Davies, 1987).

The previous suggestion that photosynthesis and the transport of the photoassimilates to the roots in the coffee seedlings were affected by *P. coffeae* is confirmed by the data in Figure 2D. The decrease of radioactivity in the roots (Figure 1B and F) indicates that less sucrose was transported to the roots from the leaves and consequently there was a decrease in the specific radioactivity of sucrose. Being transported less to the roots and not being diluted by the sucrose

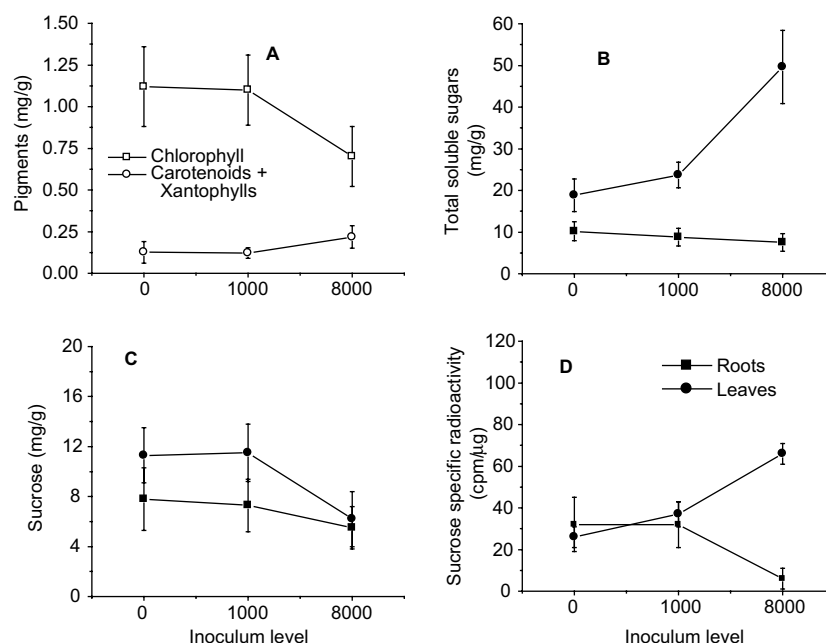


Figure 2. The effect of *P. coffeae* on pigment contents (A), total soluble sugars (B), sucrose (C), and specific radioactivity of sucrose (D) in coffee seedlings of the assay 2. Bars indicated standard deviation. Means of five (assay 1) and eight (assay 2) replicates.

formed after the plastic bags containing $^{14}\text{CO}_2$ had been removed, because of low photosynthesis, the specific radioactivity in the leaves can be expected to increase in infested plants.

In both assays, on most occasions, significant alterations were observed only following infestation with 8000 nematodes. This is in agreement with a previous study with coffee seedlings infested with *P. coffeae* where it was demonstrated that photosynthesis was decreased at inoculation levels higher than 2250 (Kubo et al., 2003).

Therefore, we conclude from our data obtained with coffee seedlings that the effects of *P. coffeae* on the carbon assimilation and partition are quite distinct from those with root knot nematodes. In contrast to these nematodes, where the feeding sites are regarded as strong metabolic sinks even leading to increase of photosynthesis in the beginning of infestation, the root lesion nematodes caused a rapid detrimental effect on carbon fixation and photoassimilates distribution in the plant due to direct damage of the roots. The same reason seems to explain the decreased uptake of nitrate and ammonium in coffee infested with *P. coffeae* (Vaast et al., 1998).

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References

- Bird AF and Loveys BR (1975) The incorporation of photosynthates by *Meloidogyne javanica*. *Journal of Nematology* 7: 111–113.
- Campos VP, Sivapalan P and Gnanapragasam NC (1990) Nematode parasites of coffee, cocoa and tea. In: Luc M, Sikora RA and Bridge J (eds) *Plant Parasitic Nematodes in Subtropical and Tropical Agriculture* (pp 387–430) CAB International, Wallingford, UK.
- Carneiro RG, Mazzafera P and Ferraz LCCB (2000) Carbon partitioning in soybean infected with *Meloidogyne incognita* and *M. javanica*. *Journal of Nematology* 31: 348–355.
- Chaplin MF (1986) Monosaccharides. In: Chaplin, MF and Kennedy JF (eds) *Carbohydrate Analysis: A Practical Approach* (pp 1–36) IRL Press, Oxford, Washington DC.
- Davies E (1987) Plant Responses to wounding. In: Davies DD (ed) *Physiology of Metabolism* (pp 243–264) Academic Press, New York.
- Dubois MK, Giller KA, Hamilton JK, Ribers PA and Smith T (1956) Colorimetric method for determination of sugars

- and related substances. *Analytical Chemistry* 28: 360–365.
- Duncan LW, Inserra RN, Thomas WK, Dunn D, Mustika I, Frisse LM, Mendes ML, Morris K and Kaplan DT (1999) Molecular and morphological analyses of isolates of *Pratylenchus coffeae* and closely related species. *Nematropica* 29: 61–80.
- Edwards DI and Wehunt EJ (1973) Hosts of *Pratylenchus coffeae* with additions from Central American banana-producing areas. *Plant Disease Reporter* 57: 47–50.
- Hussey RS (1985) Host-parasite relationships and associated physiological changes. In: Sasser JN and Carter CC (eds), *An Advanced Treatise on Meloidogyne* Vol. 1 (pp 143–153) North Caroline State University Graphics, Raleigh.
- Inomoto MM, Oliveira CMG, Mazzafera P and Gonçalves W (1998) Effects of *Pratylenchus brachyurus* and *P. coffeae* on seedlings of *Coffea arabica*. *Journal of Nematology* 30: 362–367.
- Inserra RN, Duncan LW, Troccoli A, Dunn D, Maia dos Santos J, Kaplan D and Vovlas, N (2001) *Pratylenchus jaehni* sp. n. from citrus in Brazil and its relationship with *P. coffeae* and *P. loosi* (Nematoda: Pratylenchidae). *Nematology* 3: 653–665.
- Kubo RK, Silva RA, Tomazini MD, Oliveira CMG, Mazzafera P and Inomoto MM (2003) Patogenicidade de *Pratylenchus coffeae* em plântulas de cafeeiro cv. Mundo Novo. *Fitopatologia Brasileira* 28: 41–48.
- Lichtenthaler HK and Wellburn AR (1983) Determination of total carotenoids and chlorophylls *a* and *b* of leaf extracts in different solvents. *Biochemical Society Transactions* 11: 591–592.
- Monteiro AR and Lordello LGE (1974) Encontro do nemátode *Pratylenchus coffeae* atacando cafeeiro em São Paulo. *Revista de Agricultura* 49: 164.
- Poskuta JW, Dropkin VH and Nelson CJ (1986) Photosynthesis, photorespiration, and respiration of soybean after infection with root nematodes. *Photosynthetica* 20: 405–410.
- Riedel RM, Forter JG and Mai WF (1973) A simplified medium for monoxenic culture of *Pratylenchus penetrans* and *Ditylenchus dipsaci*. *Journal of Nematology* 5: 71–72.
- Silva RA and Inomoto MM (2002) Host-range characterization of two *Pratylenchus coffeae* from Brazil. *Journal of Nematology* 34: 135–139.
- Sturm A (1999) Invertases: primary structures, functions, and roles in plant development and sucrose partitioning. *Plant Physiology* 121: 1–7.
- Southey JF (1986) *Laboratory Methods for Work with Plant and Soil Nematodes* Her Majesty's Stationery Office. London.
- Vaast Ph, Caswell-Chen EP and Zasoski RJ (1998) Effects of two endoparasitic nematodes (*Pratylenchus coffeae* and *Meloidogyne konaensis*) on ammonium and nitrate uptake by Arabica coffee (*Coffea arabica* L.). *Applied Soil Ecology* 10: 171–178.
- Van Handel E (1968) Direct microdetermination of sucrose. *Analytical Biochemistry* 22: 280–283.